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13723-002001

U.S. APPLICATION NO. (If Known, see 37 CFR 1.5)

10/009151

INTERNATIONAL APPLICATION NO.
PCT/JP00/03413INTERNATIONAL FILING DATE
26 May 2000PRIORITY DATE CLAIMED
02 June 1999TITLE OF INVENTION
STABILIZED DENATURED LIPOPROTEIN AND PROCESS FOR PRODUCING THE SAMEAPPLICANT(S) FOR DO/EO/US
Takashi Shigematsu, Kyoko Shimamura, Junji Kimura, Hiroaki Kohno and Nobuyuki Sueshige

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This is an express request to promptly begin national examination procedures (35 U.S.C. 371(f)).
4. ☐ The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 16 below concern other documents or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A FIRST preliminary amendment.
 - ☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☐ Other items or information:



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U.S. APPLICATION NO. (IF KNOWN) 10/009151	INTERNATIONAL APPLICATION NO. PCT/JP00/03413	ATTORNEY'S DOCKET NUMBER 13723-002001
17. <input checked="" type="checkbox"/> The following fees are submitted: Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1040 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$890 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$740 International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$710 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT =</div>		CALCULATIONS PTO USE ONLY <div style="border: 1px solid black; height: 150px; width: 100%;"></div>
Surcharge of \$130 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).		<div style="border: 1px solid black; width: 100%; height: 20px;"></div>
Claims	Number Filed	Number Extra
Total Claims	21 - 20 =	1
Independent Claims	3 - 3 =	0
MULTIPLE DEPENDENT CLAIMS(S) (if applicable)		+ \$280
TOTAL OF ABOVE CALCULATIONS =		\$1,008.00
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.		<div style="border: 1px solid black; width: 100%; height: 20px;"></div>
SUBTOTAL =		\$1,008.00
Processing fee of \$130 for furnishing the English Translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).		<div style="border: 1px solid black; width: 100%; height: 20px;"></div>
TOTAL NATIONAL FEE =		\$1,008.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) \$40.00 per property +		<div style="border: 1px solid black; width: 100%; height: 20px;"></div>
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a. <input checked="" type="checkbox"/> A check in the amount of \$1,008.00 to cover the above fees is enclosed. b. <input type="checkbox"/> Please charge my Deposit Account No. 06-1050 in the amount of \$0.00 to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 06-1050. A duplicate copy of this sheet is enclosed.		
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.		
SEND ALL CORRESPONDENCE TO:		
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		NAME: Y. Rocky Tsao
		REGISTRATION NUMBER: 34,053

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Takashi Shigematsu et al.
Serial No. : Unassigned
Filed : Herewith
Title : TRANSMISSION

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PRELIMINARY AMENDMENT

Prior to examination, please amend the application as follows:

In the claims:

Amend claims 3-8, 12-13, and 15 as follows:

--3. (Amended) A method according to claim 1, wherein said lipoprotein is at least one selected from the group consisting of chylomicrons, VLDL, LDL, Lp(a), HDL2, and HDL3.-

--4. (Amended) A method according to claim 1, wherein said denatured lipoprotein is obtained by oxidizing lipoprotein in the presence of a metal ion.--

--5. (Amended) A method according to claim 4, wherein said metal ion is a copper ion, an iron ion, or a mixture thereof.--

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Serial No. : Unassigned
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--6. (Amended) A method according to claim 1, wherein said denatured lipoprotein is obtained by acetylating lipoprotein.--

--7. (Amended) A method according to claim 1, wherein said denatured lipoprotein is obtained by incorporating aldehyde in lipoprotein by the use of malondialdehyde.--

--8. (Amended) A method according to claim 1, which further comprises a step of adding a stabilizing agent.--

--12. (Amended) A method according to claim 10, wherein said denatured lipoprotein is an oxidized lipoprotein or malondialdehyde-conjugated lipoprotein.--

--13. (Amended) A method according to claim 10, wherein said denatured lipoprotein is capable of reacting with a DLH3 antibody which is yielded by hybridoma cell line, mouse-mouse hybridoma FOH1a/DLD3 (Deposit No. FERM BP-7171).--

--15. (Amended) Stabilized denatured lipoprotein produced by a method set forth in claim 1.--

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REMARKS

All amendments have been made to remove multiple dependency while conserving the claimed subject matter. No new matter has been introduced.

Attached is a marked-up version of the changes being made by the current amendment.

Claims 1-21 are now pending. Applicant submits that all of the claims are now in condition for examination, which action is requested. Please apply any charges or credits to Deposit Account No. 06-1050 referencing attorney docket no. 13723-002001.

Respectfully submitted,

Date: 11-30-01

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Version with markings to show changes made

In the claims:

Claims 3-8, 12-13, and 15 have been amended as follows:

3. (Amended) A method according to claim 1 [or 2], wherein said lipoprotein is at least one selected from the group consisting of chylomicrons, VLDL, LDL, Lp(a), HDL2, and HDL3.
4. (Amended) A method according to [any one of claims 1 - 3] claim 1, wherein said denatured lipoprotein is obtained by oxidizing lipoprotein in the presence of a metal ion.
5. (Amended) A method according to [claims] claim 4, wherein said metal ion is a copper ion, an iron ion, or a mixture thereof.
6. (Amended) A method according to [any one of claims 1 - 3] claim 1, wherein said denatured lipoprotein is obtained by acetylating lipoprotein.
7. (Amended) A method according to [any one of claims 1 - 3] claim 1, wherein said denatured lipoprotein is obtained by incorporating aldehyde in lipoprotein by the use of malondialdehyde.
8. (Amended) A method according to [any one of claims 1 - 7] claim 1, which further comprises a step of adding a stabilizing agent.
12. (Amended) A method according to claim 10 [or 11], wherein said denatured lipoprotein is an oxidized lipoprotein or malondialdehyde-conjugated lipoprotein.
13. (Amended) A method according to claim 10 [or 11], wherein said denatured lipoprotein is capable of reacting with a DLH3 antibody which is yielded by hybridoma cell line,

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mouse-mouse hybridoma FOH1a/DLD3 (Deposit No. FERM BP-7171).

15. (Amended) Stabilized denatured lipoprotein produced by a method set forth in [any one of claims 1 - 9 or in claim 14] claim 1.

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10/PATS

DESCRIPTION**STABILIZED DENATURED LIPOPROTEIN
AND METHOD FOR PRODUCING THEREOF****Technical Field**

5 The present invention relates to stabilized denatured lipoprotein and a method for the production thereof. More particularly, the present invention relates to denatured lipoprotein excelling in stability of prolonged preservation and obtained by freeze-drying denatured lipoprotein resulting
10 from denaturing lipoprotein and a method for the production thereof.

 More particularly, the present invention relates to a method for the production of denatured lipoprotein which comprises denaturing lipoprotein by subjecting a solution
15 containing this lipoprotein to a step including at least one freezing operation, the denatured lipoprotein excelling in stability of prolonged preservation obtained by further freeze-drying the denatured lipoprotein produced in the manner described above, and a method for the production
20 thereof.

 The denatured lipoprotein strongly suggests the association thereof with various diseases of the circulatory system including such diseases of the coronary artery system as cardiac infarction and stenocardia, such diseases of the
25 cerabral arteries as cerebral infarction and cerebravascular dementia, such diseases of the renal arteries as nephropathy and diabetic nephropathy, and such diseases of the peripheral artery system as obstruction of peripheral arteries. The standard substances for determining mass of denatured
30 lipoprotein and the various experimental reagents for the investigation of physiological role and physiological activity of the denatured lipoprotein, therefore, constitute

themselves very important substances that affect the results of the research efforts. The denatured lipoprotein which has been stabilized in the manner described above, therefore, is useful as a standard substance in a method for determining
5 denatured lipoprotein content in a blood component as by causing the denatured lipoprotein to contact an antibody capable of recognizing the denatured lipoprotein and determining the reactivity of the antibody with a sample and as a varying experimental reagent for investigating the
10 physiological role and physiological activity of the denatured lipoprotein.

Background Art

In various diseases of the circulatory system including such diseases of the coronary artery system as cardiac
15 infarction and stenocardia, such diseases of the cerebral arteries as cerebral infarction and cerebrovascular dementia, such diseases of the renal arteries as nephropathy and diabetic nephropathy, and such diseases of the peripheral artery system as obstruction of peripheral arteries, it is strongly
20 suggested that the lipid in the blood serum plays an important role. A huge amount of expenses are being paid today by the public health system for the medical treatments using serum lipid depressants, particularly cholesterol depressants. Recent studies have yielded a report that comparison between
25 a group of patients of such diseases and a group of healthy persons reveals no noticeable difference in the absolute amount of serum lipid between the two groups and that rather the amount of oxidized LDL, a denatured product of low-density lipoprotein (LDL), present in the blood serum is distinctly
30 different between the two groups (Toshima, S. Et al. (1996) Circulation, 94, Suppl. I: 1288). The relationship between the oxidized lipoprotein, one of the denatured lipoproteins,

and the advance of atherosclerotic lesion has been pointed out by Steinberg et al. (Steinberg, D., Parthasarathy, S. Carew, T. E., Khoo, J. C., and Witztum, J. L., (1989) N. Engl. J. Med., 320: 915) for example. In recent years, therefore, various methods for determination using the denatured lipoprotein have been under development (Patent publication JP-A-08-304,395 and Patent publication JP-A-09-288,106, for example) and the test for investigating the physiological role of the denatured lipoprotein has been gaining in importance.

The fact that it is difficult to obtain the standard substances necessary for the determinations performed in such experiments, however, has complicated the situation. For the purpose of investigating the physiological role of denatured lipoprotein, it becomes necessary to collect a large number of blood samples denatured lipoprotein from a plurality of installations and compare them. For the comparison, it is essential that the measurements obtained in the individual tests be free from variation. No reproducibility can be ensured among these measurements unless a standard substance stable and excellent in reproducibility throughout a duration necessary for a series of relevant tests is available. When the test results are markedly varied among different persons who measure owing to the use of different standard substances, the interpretation of the physiological role of given denatured lipoprotein is so complicated as to render it impossible to obtain a fixed conclusion. This inability to obtain a standard substance which fits stable preservation has closed the way of applying the determination of the amount of denatured lipoprotein present in a given sample, for example, as a means to judge a disease exactly despite popular recognition of the physiological importance of this

determination.

Generally, in the determination of the mass of protein contained in blood serum, for example, the practice of stabilizing by some method or other what may be called a standard blood serum or a target component in an isolated state and using the product of this stabilization as a standard substance is now carried out. As regards the lipoprotein in general, a method for producing standard blood plasma or standard blood serum stable during prolonged preservation by optionally mixing lipoprotein-containing blood plasma or blood serum with such nonreducing sugar as sucrose and freeze-drying the resultant mixture till the water content reaches a level in the range of 1 - 10 mass % (Patent publication EP-A-617289) and a method for commercially producing a stable freeze-dried product by obtaining reconstructive lipoprotein from apolipoprotein and lipid and freeze-drying and stabilizing the reconstructive lipoprotein in the presence of a stabilizer such as sucrose or mannitol (Patent publication US-A-5,652,399) have been reported. The patents published in these official gazettes merely attempt to stabilize lipoprotein as a means to stabilize the lipoprotein without entailing denaturation thereof and do not disclose the stabilized denatured lipoprotein and a method for the production thereof.

A method for obtaining such denatured lipoprotein as oxidized lipoprotein, acetylated lipoprotein, or malondialdehyde-conjugated lipoprotein by isolating and purifying lipoprotein by a heretofore well-known technique for example, ultracentrifugation and then oxidizing the refined lipoprotein with such a metal ion as a copper ion, acetylating it by the reaction with acetic anhydride, or allowing it to react with malondialdehyde has been known.

The denatured lipoproteins produced by this method, however, are more unstable than the undenatured lipoprotein and unable in their denatured state to allow prolonged preservation.

In view of this state of affairs, Patent publication
5 JP-A-09-288,106 has disclosed a method for determining human oxidized lipoprotein by using as a standard what is produced by incorporating into blood plasma lipoprotein the oxide of phospholipid obtained by artificially oxidizing phospholipid. The standard substance which is used in the method described
10 above, however, is so deficient in stability of preservation as to require the individual preparation thereof prior to each use and entail a complicated procedure.

In view of such various states of affairs, the development of denatured lipoprotein capable of being stably stored for
15 a long time and a method for the production thereof has been desired for.

Therefore objects of the present invention are to provide denatured lipoprotein excelling in stability of prolonged preservation, namely, the lipoprotein does not cause any
20 discernible variation in the determinations throughout the duration of preservation and a method of the production thereof, wherein the lipoprotein is applied as a standard substance for the determination of the mass of denatured lipoprotein contained in a given blood component or as a varying
25 experimental reagent for the investigation of physiological role of denatured lipoprotein.

Disclosure of the Invention

The present inventors, as a result of a diligent study pursued with a view to attaining the object mentioned above,
30 have found that by freeze-drying such artificially denatured lipoproteins as; oxidized lipoproteins obtained by oxidizing such lipoproteins as egg yolk, milk, whole blood, blood serum,

and blood plasma severally containing lipoprotein, lipoprotein fractions partially fractionated therefrom, and lipoproteins which are fractionally purified by the ultracentrifugal separation technique, by the use of a catalyst represented by such a metal ion as a copper ion; acetylated lipoprotein obtained by acetylating the above-mentioned lipoproteins with acetic anhydride etc.; or malondialdehyde-conjugated lipoprotein obtained by reacting the above-mentioned lipoprotein with malondialdehyde etc., it is made possible to improve conspicuously such denatured lipoproteins in stability of prolonged preservation and consequently accomplish the object of the present invention. The present inventors have also found that by allowing the presence of sucrose, lactose, trehalose, bovine blood serum albumin (BSA), or human blood serum albumin (HSA) etc. as stabilizer during the course of freeze-drying, it is made possible to improve the denatured lipoprotein to a greater extent in stability of prolonged preservation and give a better solution to the problems mentioned above.

The present inventors, after a diligent study pursued with a view to accomplishing the object mentioned above, have found that by performing a process including at least one freezing operation on a solution containing lipoprotein thereby denaturing the lipoprotein contained in the solution, it is made possible to obtain a standard substance usable for the determination of a denatured protein content in a given blood sample or a varying experimental reagent usable for the investigation of physiological role or physiological activity of the denatured protein. They have further found that by freeze-drying the denatured lipoprotein obtained as described above, it is made possible to improve prominently the stability of prolonged preservation of the denatured

lipoprotein in the dried state and the stability of preservation of the denatured lipoprotein in the dried state after it has been dissolved into a solution and consequently accomplish the object of the present invention. Here again, 5 the present inventors have further found that by allowing the presence of sucrose, lactose, trehalose, bovine blood serum albumin (BSA), or human blood serum albumin (HSA) etc. as a stabilizer during the course of freeze-drying, it is made possible to improve the denatured lipoproteins to a 10 greater extent in stability of prolonged preservation and give a better solution to the problems mentioned above.

The present invention has been perfected based on such knowledge as mentioned above.

To be specific, the object mentioned above is accomplished by a method for producing stabilized denatured 15 lipoprotein by artificially denaturing lipoprotein, thereby obtaining denatured lipoprotein and freeze-drying the denatured lipoprotein, thereby stabilizing the denatured lipoprotein.

20 The object mentioned above is also accomplished by a method for producing denatured lipoprotein by performing a process including at least one freezing operation on a solution containing lipoprotein, thereby denaturing the lipoprotein contained in the solution.

25 The object is further accomplished by a method for producing stabilized denatured lipoprotein, which comprises performing a process including at least one freezing operation on a solution containing lipoprotein, thereby denaturing the lipoprotein contained in the solution and obtaining denatured 30 lipoprotein and further freeze-drying the denatured lipoprotein, thereby stabilizing the denatured lipoprotein.

Brief Description of the Drawings

Fig. 1 is a diagram showing the calibration curve produced with the oxidized LDL obtained in Example 1 of the present invention as a sample, in which the oxidized LDL is prepared by freeze-drying the LDL oxidized with copper and then
5 dissolving the dry LDL by a prescribed method.

Fig. 2 is a diagram comparing the determinations obtained from the sample prepared by preserving the oxidized LDL obtained in Example 1 of the present invention at 4°C in which the oxidized LDL is prepared by freeze-drying the LDL oxidized
10 with copper, and dissolving the LDL at stated durations by a prescribed method, and, the determinations at stated durations obtained from the sample prepared by preserving the LDL oxidized with copper at 4°C without being freeze-dried.

Fig. 3 is a diagram showing the calibration curve produced with the oxidized HDL obtained in Example 2 of the present
15 invention as a sample, in which the oxidized HDL is prepared by freeze-drying the HDL oxidized with copper and then dissolving the dried HDL by a prescribed method.

Fig. 4 is a diagram comparing the determinations obtained from the sample prepared by preserving the oxidized HDL obtained in Example 2 of the present invention at 4°C in which the oxidized HDL is prepared by freeze-drying the HDL oxidized
20 with copper, and dissolving the HDL at stated durations by a prescribed method, and, the determinations at stated durations obtained from the sample prepared by preserving the HDL oxidized with copper at 4°C without being freeze-dried.

Fig. 5 is a diagram showing the calibration curve produced with the oxidized lipoprotein a [Lp(a)] obtained in Example
3 of the present invention as a sample, in which the oxidized
30 Lp(a) is prepared by freeze-drying the LDL oxidized with copper and then dissolving the dried Lp(a) by a prescribed method.

Fig. 6 is a diagram comparing the determinations obtained

from the sample prepared by preserving the oxidized Lp(a) obtained in Example 3 of the present invention at 4°C in which the oxidized Lp(a) is prepared by freeze-drying the Lp(a) oxidized with copper, and dissolving the Lp(a) at stated durations by a prescribed method, and, the determinations at stated durations obtained from the sample prepared by preserving the Lp(a) oxidized with copper at 4°C without being freeze-dried.

Fig. 7 is a diagram illustrating the production of denatured lipoprotein by performing a process including a step of freezing on blood plasma as demonstrated in Example 4.

Fig. 8 is a diagram for comparing the determinations (absorbance) obtained in Example 5 respectively of the samples of standard freeze-denatured human blood serum prepared in Example 5 (2) and the samples of standard oxidized LDL prepared in Example 1 (3) after the samples had been dissolved and rated for stability of preservation according to the ELISA method.

Fig. 9 is a diagram illustrating the production of denatured lipoprotein by performing a process including a freezing step on human LDL in Example 6.

Fig. 10 is a diagram for comparing the determinations (absorbance) obtained in Example 6 respectively of the samples of standard freeze-denatured LDL prepared in Example 6 (2) and the samples of standard oxidized LDL prepared in Example 1 (3) after the samples had been dissolved and rated for stability of preservation according to the ELISA method.

Best Mode of Embodying the Invention

Now, the present invention will be described in detail below.

The first aspect of the present invention consists in

providing a method for producing stabilized denatured lipoprotein, comprising a step of artificially denaturing lipoprotein, thereby obtaining denatured lipoprotein and a step of freeze-drying said denatured lipoprotein, thereby
5 stabilizing said denatured lipoprotein.

The term "denatured lipoprotein" as used in this specification means both the denatured lipoprotein which has undergone such a chemical change as oxidized lipoprotein, acetylated lipoprotein and malondialdehyde-conjugated
10 lipoprotein obtained by action of malondialdehyde etc., and the denatured lipoprotein which has undergone such a structural change as coagulation or change in three-dimensional structure and also includes such kinds of denatured lipoprotein as identified by a change in electric
15 charge, a change in molecular weight, a change in affinity for an in vivo receptor, and a change in ability to form linkage with denatured lipoprotein specific antibody (J. Biol. Chem. 1994, 269: 15274-15279 and Patent publication JP-A-07-238,098) represented by the antibody yielded by
20 FOH1a/DLH3 (Deposit No.: FERM BP-7171) comparing to an undenatured lipoprotein. The expression "change in ability to form linkage with denatured lipoprotein specific antibody represented by the antibody yielded by FOH1a/DLH3" as mentioned above refers to the difference between the amount
25 of signal obtained from an undenatured lipoprotein antigen and the amount of signal obtained from denatured lipoprotein antigen in an operation performed by transforming the undenatured lipoprotein and the denatured lipoprotein conform to the present invention each as an antigen into an
30 insolubilized solid phase, causing a relevant antibody to react with the solidified antigen, and detecting the amount of the antibody consequently bound to the solidified antigen

as converted to the enzyme content by the use of an enzyme standardized antibody possessing specificity to the antibody.

The lipoprotein to be used in the present invention may originate in any of the living organisms. As concrete examples of the lipoprotein, the lipoproteins which originate in such mammals as man, cow, horse, goat, sheep, rabbit, dog, and guinea pig; such birds as fowl and quail; such fishes as salmon and herring; and such microorganisms as bacteria and mycetes may be cited. Further, as concrete examples of the lipoprotein to be used in the present invention, structural lipoproteins which occur in such biomembranes as cell membrane, mitochondrial cell, myelinic structure membrane, and bacterial cell membrane; soluble lipoproteins which occur in blood plasma, egg yolk, and milk; and such lipoprotein fractions obtained by fractionating these lipoproteins mentioned above by the ultracentrifugation technique to chylomicron, very low-density lipoprotein (VLDL), low-density lipoprotein (LDL), lipoprotein X, intermediate density lipoprotein (IDL), lipoprotein a [Lp(a)], high-density lipoproteins (HDL) like HDL2 and HDL3, and very high-density lipoprotein (VHDL) may be cited. These lipoproteins may be used either singly or in the form of a combination of two or more members. Of these lipoproteins, those of human origins are used preferably, chylomicron, VLDL, LDL, Lp(a), HDL2 or HDL3 originating in human blood plasma and blood serum, and mixtures thereof are used more preferably, and LDLs originating in human blood plasma and blood serum are used most preferably.

The human lipoprotein which is typically used in the present invention is prepared by separating a fraction with a prescribed specific gravity from human blood serum by such a universally known technique as centrifugal sedimentation

or ultracentrifugation and purifying this fraction by such a heretofore known technique as dialysis or desalination. As concrete examples of the method for the preparation of the low-density lipoprotein (LDL), the following methods (1)

5 - (3) may be cited.

(1) To 20-30mL of normal human blood serum, ethylenediaminesodiumtetraacetate (EDTA-2Na) is added until a final concentration of 1 mmol/L. The resultant mixture is made to add NaBr with adjusting to a specific gravity of 1.000.
10 The produced mixture is dispensed into centrifugal tubes, thereby obtained the portions of the mixture in the respective tubes. Buffer solutions is adjusted with NaBr to varied levels of specific gravity of 1.15, 1.063, 1.019, and 1.006, and superposed respectively on the portions of mixture in sequence.
15 Then the tubes contained the mixtures and the buffers are together centrifuged ($120,000\times g$) at 4°C for 24 hours. The fractions consequently formed are sequentially separated from the upper to the down and measured for specific gravity with a refractometer to collect fractions having a specific gravity
20 in the range of 1.019 - 1.063 as LDL fractions. The LDL fractions which have been obtained as described above are dialyzed, immediately after collection, with PBS [phosphate buffer of 10 mmol/L and NaCl (pH 7.4) of 140 mmol/L] including 0.25 mmol/L EDTA (Patent publication JP-A-07-238,098,
25 paragraph No. 0040).

(2) To the heparinized human blood plasma, EDTA is added until a final concentration of 0.25 mmol/L. The blood plasma thus prepared is dispensed in a unit volume of 0.75 mL into centrifugation tubes (1 - 4mL). The portions of the blood
30 plasma in the respective tubes and 0.15 mol/L NaCl of 250 μL containing 0.3 mmol/L EDTA and superposed in a unit volume of 250 μL on the portions are together centrifuged with

185,000×g at 10°C for 2.5 hours. 150 μL of the upper layer is discarded. 750 μL of the lower layer is taken and then made to add 150 μL of KBr solution (50 w/v %) until a specific gravity of 1.063. The blood plasma samples with the adjusted
5 specific gravity are moved to the bottoms of the centrifugation tubes (1 - 4mL) and centrifuged with 244,000×g at 10°C for 16 hours. The orange band in the upper layer (about 100 - 150μL) is attentively collected and dialyzed against PBS containing 0.25 mmol/L EDTA at 4°C for 6 hours (3 liters of
10 PBS replaced twice by two hours interval) (Patent publication JP-A-08-304,395, paragraph No. 0050); or

(3) From the human blood plasma obtained with EDTA as an anti-coagulating agent, the portion having specific gravity 1.019 - 1.063 is collected as a LDL fraction by the
15 centrifugation technique. The LDL fraction, after having the purity thereof verified by the formation of a single sharp band on analysis by the agarose electrophoresis, is thoroughly dialyzed against a PBS solution (pH 7.4) containing 0.25 mmol/L EDTA (Patent publication JP-A-09-288,106, paragraph
20 No. 0062).

Then, as one concrete example of the method for preparing lipoprotein a [Lp(a)], for instance, the following method may be cited: To the heparinized human blood plasma, EDTA is added until a final concentration of 0.25 mmol/L. The blood
25 plasma sample thus prepared and 250 μL of 0.15 mol/L NaCl containing 0.3 mmol/L EDTA and superposed on the sample are together centrifuged with 105,000×g at 8°C for 20 hours. The upper layer is discarded. In the lower layer, KBr pulverized by a mortar in advance is dissolved until a final specific
30 gravity of 1.125 while avoiding formation of bubbles. The resultant solution is centrifuged with 105,000×g at 8°C for 20 hours. The orange band in the upper layer is collected

attentively and then subjected to gel filtration with Biogel A-5m (Bio-rad Corp.) with 1 mol/L NaCl, 2 mmol/L EDTA, and 10 mmol/L phosphate buffer as a developing solvent. The fractions consequently obtained are each assayed with a Lp(a) measuring kit (produced by Thermo K.K.) to collect the Lp(a) fraction. This fraction is treated with lysine sepharose 4B (Farmacia Corp.). The fraction adsorbed thereon is eluted with a buffer containing 0.2 mol/L ϵ -aminocaproic acid and dialyzed against PBS containing 0.25 mmol/L EDTA (Patent publication JP-A-08-304,395, paragraph No. 0052).

The method for artificially denaturing lipoprotein in the present invention does not need to be particularly discriminated but may be properly selected from the known methods. As concrete examples of the method for denaturing human lipoprotein, for instance, a method which consists in oxidizing the human lipoprotein in the presence of a metal ion, a method which consists in acetylating the human lipoprotein, a method which consists in incorporating aldehyde in the human lipoprotein by the use of malondialdehyde, and a method which consists in adding a solution dissolved a compound obtained by artificially oxidizing such lipoprotein as 1-palmitoyl-2-(9-oxononanoyl)-glycerol-3-phosphocholine and 1-palmitoyl-2-(5-oxovaleroyl)-glycerol-3-phosphocholine in a proper solvent like DMSO to the human lipoprotein may be cited. In the methods enumerated above, the method which consists in oxidizing the human lipoprotein in the presence of a metal ion, the method which consists in acetylating the human lipoprotein, and the method which consists in having aldehyde incorporated in the human lipoprotein by the use of malondialdehyde are used favorably.

Now, the three preferred methods mentioned above will be described in detail below as typical modes of embodiment of the invention.

Firstly, the mode of embodiment which consists in
5 oxidizing the human lipoprotein in the presence of a metal ion will be described in detail below. One example of the mode of embodiment mentioned above will be described below: The human lipoprotein (fraction) which has been prepared as described above is deprived of EDTA by dialyzing against a
10 buffer containing no EDTA [for example, PBS (pH 7.4)] and adjusted to a prescribed protein content (50 - 2000 $\mu\text{g/mL}$, preferably 100 - 500 $\mu\text{g/mL}$). It is made to add copper sulfate (CuSO_4) until a prescribed concentration and then left reacting therewith at a temperature in the approximate range of 36
15 - 38°C.

As concrete examples of the metal ion to be used in the mode of embodiment mentioned above, copper ions originating in copper (II) fluoridedihydrate, copper (II) bromide, copper (II) oxide, copper (II) hydroxide, copper (II) sulfate, copper
20 (II) sulfate pentahydrate, copper (I) selenide, copper (II) selenide, copper (II) selenide pentahydrate, copper (II) hexafluorosilicate tetrahydrate, copper (II) acetate monohydrate, copper (II) tetraammine sulfate monohydrate, and copper (II) bis(ethylenediamine) sulfate dihydrate; iron
25 ions originating in iron (II) chloride, iron (II) bromide hexahydrate, iron (II) nitrate hexahydrate, iron (II) thiocyanate trihydrate, iron (II) acetate tetrahydrate, iron (III) oxalate pentahydrate, iron (II) ammonium sulfate hexahydrate, iron (III) potassium sulfate dodecahydrate, iron
30 (II) ammonium sulfate dodecahydrate, and iron sulfate; metal ions of hemoglobin (Hb), transferrin (Tf), and lactoferrin (Lf) and mixtures thereof may be cited. Among other metal

ions enumerated above, the copper ions originating in copper (II) sulfate and copper (II) sulfate pentahydrate and mixtures thereof are preferably used. These metal ions may be used either singly or in the form of a combination of two or more members.

The amount of the metal ion to be used in the mode of embodiment described above does not need to be particularly discriminated but is only required to permit thorough oxidation of the human lipoprotein. It may be such that the concentration of the metal ion will be in the range of 10 - 200 $\mu\text{mols/L}$, preferably in the range of 25 - 100 $\mu\text{mols/L}$, based on 1 g of the human lipoprotein to be oxidized.

In the mode of embodiment described above, if the reaction time is unduly short, the shortage will be at a disadvantage in preventing the denaturation (oxidation) of lipoprotein from being effected sufficiently. Conversely, if the reaction time is unduly long, the excess will be at a disadvantage in forcing the lipoprotein itself to decompose excessively and fail into the loss of the antigenicity of the apoprotein, for example. Though the reaction time cannot be generally defined because it varies with the amount of the residual EDTA and the total amount of the solution, it is in the range of 1 - 24 hours, preferably in the range of 1-24 hours, preferably 2-4 hours, at temperatures of about 36°C - 38°C in the case of the lipoprotein (fraction) prepared in the mode of embodiment described above.

Secondly, the mode of embodying the acetylation of the human lipoprotein will be described in detail below. One example of this mode of embodiment will be described below: The solution of the human lipoprotein (fraction) prepared in a prescribed protein concentration (about 500 - 2000 $\mu\text{g/mL}$, preferably 500-1000 $\mu\text{g/mL}$) as described above and saturated

sodium acetate etc. added thereto in a volume equal thereto are stirred together at 0 °C - 4°C for 1 - 2 hours. Then, the resultant mixture and acetic anhydride added thereto in a volume in the range of 0.25 - 4 µL (namely 0.5 - 2 µL/mg, based on the amount of the human lipoprotein), preferably in the range of 0.4 - 2.4 µL (namely 0.8 - 1.2 µL/mg, based on the amount of the human lipoprotein) are stirred together at 0 °C - 4°C for 60 - 120 minutes. Then, the produced mixture is dialyzed thoroughly against PBS (pH 7.4) containing 0.25 mmol/L of EDTA of 500-1000 times as large in volume of the produced mixture, at 2 °C - 8°C, two or three times for not less than two hours/each.

Thirdly, the mode of embodying the incorporation of aldehyde in the human lipoprotein by the use of malondialdehyde will be described in detail below. One example of the mode of embodiment mentioned above will be described below: The solution of the human lipoprotein (fraction) prepared as described above in a prescribed protein concentration (about 0.25 - 1 mg/mL, preferably 0.25 - 0.5 mg/mL) in a 0.1 mol/L phosphate buffer (pH 6.5) and a malondialdehyde solution (obtained by thermally hydrolyzing 1 mol/L of malondialdehyde bisdimethyl acetal at 100°C for five minutes in the presence of 0.1 mol/L of hydrochloric acid) added thereto in a volume in the range of 0.625 - 10 µL (namely 2.5 - 10 µL/mg, based on the amount of the human lipoprotein), preferably in the range of 1 - 6 µL (namely 4 - 6 µL/mg, based on the amount of the human lipoprotein) are left reacting with each other at 30°C - 40°C for 2 - 4 hours. The resultant reaction solution is dialyzed thoroughly against PBS (pH 7.4) containing 0.25 mmol/L of EDTA of 500-1000 times as large in volume of the produced mixture, at 2°C - 8°C, two or three times for not less than two hours/each.

The method contemplated by the present invention essentially requires including a step for freeze-drying the denatured lipoprotein obtained as described above for the purpose of stabilizing the product. In the present
5 specification, the term "freeze-drying" is used in the meaning as used in the relevant field. Specifically, it means to freeze a given sample, decompress the sample as kept in a frozen state, and deprive the sample of water and a subliming component until it dries. The conditions for the
10 freeze-drying in the present invention do not need to be particularly discriminated but are only required to be capable of stabilizing the denatured lipoprotein. The freeze-drying is generally effected at a temperature in the range of -80 °C - 20°C, preferably in the range of -80 °C - 15°C, under a
15 pressure in the range of 0.667 - 13.33 Pa, preferably in the range of 0.667 - 1.333 Pa for a period in the range of 12 - 72 hours, preferably in the range of 24 - 72 hours. By this step of freeze-drying, the water content in the freeze-dried product contained the denatured lipoprotein is generally made
20 to fall to not more than 10 mass %, preferably to not more than 1 mass %.

In the present invention, the step of freeze-drying is preferred to proceed in the presence of a stabilizing agent. The stabilizing agent to be used in the mode of embodiment
25 described above is the same stabilizing agent as used generally in the relevant field. As concrete examples of the stabilizing agent, saccharides such as sucrose, lactose, and trehalose; and proteins such as bovine blood serum albumin (BSA) and human blood serum albumin (HSA) may be cited. In these
30 substances, sucrose, lactose, trehalose, bovine blood serum albumin (BSA), and human blood serum albumin (HSA) are preferably used as stabilizing agents. Incidentally, the

stabilizing agents enumerated above may be used either singly or in the form of a mixture of two or more members. Though the amount of the stabilizing agent to be used does not need to be particularly restricted but is only required to be capable of stabilizing the denatured lipoprotein, it is generally
5 in the range of 1 - 20 mass %, preferably in the range of 2 - 5 mass %.

In the present invention, the timing of the addition of a stabilizing agent during the course of the freeze-drying
10 which is expected to proceed in the presence of the stabilizing agent does not need to be particularly restricted. The addition of the stabilizing agent preferably precedes the step of freeze-drying and particularly preferably intervenes between the step for denaturing and the step for freeze-drying.
15 Further, the step for freeze-drying does not need to be particularly followed by any step for depriving the frozen lipoprotein of the stabilizing agent. In consideration of the stability of the preservation of the denatured lipoprotein, the continued presence of the stabilizing agent while the
20 freeze-dried state is retained proves more preferable than not.

The second aspect of the present invention consists in providing a method for the production of denatured lipoprotein, comprising a step of performing a process including at least
25 one freezing operation on a solution containing lipoprotein thereby denaturing the lipoprotein contained in said solution. The third aspect of the present invention consists in providing a method for producing stabilized denatured lipoprotein, which comprises further freeze-drying the denatured
30 lipoprotein obtained as described above, thereby stabilizing the denatured lipoprotein. The present inventors have found that the denatured lipoprotein obtained by subjecting the

solution containing lipoprotein to a process including at least one freezing operation thereby denaturing the lipoprotein contained in the solution is also usable as a standard substance for the purpose of determination of the mass of denatured lipoprotein in blood and as a reagent for investigating the physiological role and the physiological activity of the denatured lipoprotein and that when the denatured lipoprotein obtained as described above is further freeze-dried, the denatured lipoprotein excels in stability of prolonged preservation in the dried state and the denatured lipoprotein in the dried state, on being dissolved in a solution, enjoys stability of preservation. The methods of the aspects mentioned above have been conceived based on this knowledge.

The method of the second or the third aspect of the present invention essentially requires to include a step for freezing the solution containing lipoprotein. The term "lipoprotein" as used in the second or the third aspect of the present invention has the same definition as in the first aspect of the invention described above. As concrete examples of the solution containing lipoprotein, blood serum, blood plasma, and chylomicrons and such lipoprotein fractions as very low-density lipoprotein (VLDL), low-density lipoprotein (LDL), lipoprotein X, intermediate density lipoprotein (IDL), lipoprotein a [Lp(a)], high-density lipoproteins (HDL) like HDL2 and HDL3, and very high-density lipoprotein (VHDL) may be cited. In the solutions enumerated above, blood serum, blood plasma, chylomicrons, VLDL, LDL, Lp(a), HDL2, HDL3, and mixtures thereof are used more preferably and human blood plasma, human blood serum, and LSLs originating in the human blood plasma and human blood serum are used most preferably as solutions containing lipoprotein.

The method according to the second aspect of the present

invention essentially requires to subject the solution containing lipoprotein to a process including a step for freezing. In the present specification, the term "process including at least one freezing operation" means a process which includes at least one step of freezing part or the whole of the water component in the lipoprotein contained in the solution or in the environment substantially encircling the lipoprotein. The denatured lipoprotein which is obtained by carrying out the process including the step for freezing as contemplated by the present invention may be any of the denatured lipoproteins enumerated in the description of the first aspect of the present invention. Preferably, the denatured lipoprotein which is obtained by carrying out the process including the step for freezing according to the present invention is oxidized lipoprotein, lipoprotein obtained by incorporation of aldehyde, or lipoprotein which reacts with the antibody yielded by a hybridoma cell line FOH1a/DLH3 (Deposit No.: FERMBP-7171) (occasionally referred to simply as "DLH3 antibody"). The hybridoma cell line FOH1a/DLH3 which is used in the method described above was deposited on February 17, 1994 with Deposit No. FERM P-14153 in National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology of the Ministry of International Trade and Industry located at 1-1-3, Higashi, Tsukuba-shi, Ibaraki-ken, Japan, under the designation of "Mouse-Mouse hybridoma FOH1a/DLH3," which deposit was switched on May 26, 2000 to the deposit based on the Budapest Treaty and has been stored at the institute with Deposit No. FERM BP-7171.

The conditions for the process including the step for freezing which is performed on the solution containing lipoprotein with a view to denaturing the lipoprotein in the

mode of embodiment mentioned above do not need to be particularly restricted but are only required to be capable of denaturing the lipoprotein contained in the solution. Further, the process including the step for freezing cannot

5 be generally restricted because the effect of freezing is greatly swayed by the environment substantially encircling the lipoprotein during the course of the freezing. The conditions for the freezing in the process including the step for freezing, for example, are expected to permit a procedure

10 which comprises lowering the temperature at a temperature decreasing speed in the range of $0.01^{\circ}\text{C} - 10^{\circ}\text{C}/\text{min}$, preferably in the range of $0.01^{\circ}\text{C} - 1^{\circ}\text{C}/\text{min}$ until it reaches a level in the range of $0^{\circ}\text{C} - -196^{\circ}\text{C}$, preferably in the range of $-5^{\circ}\text{C} - -85^{\circ}\text{C}$ and freezes and then retaining a prescribed

15 temperature for a period in the range of 0 - 36 hours, preferably in the range of 0 - 16 hours. In the second mode of embodiment, the process including the step for freezing for the purpose of denaturing lipoprotein is required to be performed at least once and, when necessary, may be repeated. When the process

20 including the step for freezing is repeated, the number of repetitions is properly in the range of 1 - 10, preferably in the range of 1 - 4. In the present invention, the contents of the repeating processes may be the same or different at each step for freezing. The conditions for the step for

25 freezing in each of the processes may be the same or different. If the number of repetitions of the step for freeze-drying exceeds 10, the excess will be at an economic disadvantage in inducing attenuation of the effect of denaturation of the lipoprotein.

30 The process including the step for freezing which aims to denature the lipoprotein in the second aspect of the invention described above may incorporate therein a step for

melting after the step for freezing. When the second aspect of the invention incorporates the step for melting in the process including the step for freezing, the conditions for the freezing do not need to be particularly restricted. The melting is attained by elevating the temperature at a temperature increasing speed in the range of $0.01^{\circ}\text{C} - 10^{\circ}\text{C}/\text{min.}$, preferably in the range of $0.1^{\circ}\text{C} - 10^{\circ}\text{C}/\text{min.}$ until it reaches a level in the range of $0^{\circ}\text{C} - 42^{\circ}\text{C}$, preferably in the range of $0^{\circ}\text{C} - 37^{\circ}\text{C}$.

The process including the step for freezing for the purpose of denaturing lipoprotein according to the second aspect of the invention described above may be such a kind of process including a step for drying in the course of the process. The conditions for the process including the step for drying after the step for freezing do not need to be particularly restricted. The drying is carried out, for example, at a temperature in the range of $-80^{\circ}\text{C} - 20^{\circ}\text{C}$, preferably in the range of $-80^{\circ}\text{C} - 15^{\circ}\text{C}$, under a pressure in the range of $0.6 - 13\text{ Pa}$, preferably in the range of $0.6 - 1.3\text{ Pa}$, for a period in the range of $12 - 72\text{ hours}$, preferably in the range of $24 - 72\text{ hours}$.

The process including the step for freezing for the purpose of denaturing lipoprotein according to the second aspect of the invention described above may be such a kind of process as comprises freeze-drying a solution containing the lipoprotein, dissolving the resultant dried solution in a solvent, and subsequently again freeze-drying the produced solution. The step for freeze-drying involved in this case has the same definition as in the first aspect of the invention. The solvent for dissolving the product obtained by the first freeze-drying does not need to be particularly restricted but is only required to be capable of dissolving the dried

product. As concrete examples of the solvent, water, deionized water, and distilled water may be cited.

The method according to the third aspect of the present invention essentially requires further to freeze dry the
5 denatured lipoprotein obtained by the method of the second aspect of the present invention described above and consequently stabilize the denatured lipoprotein. The step for freeze-drying according to the aspect described above has the same definition as in the first aspect with the
10 exception of the timing of the addition of the stabilizing agent. To be more specific, the stabilizing agent varies in kind with the contents of the process including the step for freezing in the second aspect. When the process including the step for freezing is performed on the solution containing
15 denatured lipoprotein, for example, the addition of the stabilizing agent may precede the process including the step for freezing or intervene between the step for freezing which is aimed at denaturing lipoprotein and the step for freeze-drying which is aimed at stabilizing the denatured
20 lipoprotein. When the solution containing the denatured lipoprotein is to be frozen, the stabilizing agent is preferred to be added in advance of the process including the step for freezing.

The fourth aspect of the present invention consists in
25 providing stabilized denatured lipoprotein that is produced by the first or the third aspect of the invention described above.

The denatured lipoprotein and the stabilized denatured lipoprotein which are so produced excel in stability of
30 prolonged preservation and, therefore, prove useful as a standard substance used in a method for determining denatured lipoprotein contained in a blood component by causing a given

sample to contact an antibody capable of recognizing denatured lipoprotein and measuring the reactivity of the antibody with the sample or as a varying experimental reagent for investigating the physiological role or physiological activity of denatured lipoprotein.

The fifth aspect of the present invention, therefore, consists in providing a method for the determination of denatured lipoprotein by the use of stabilized denatured lipoprotein produced in accordance with the first or the third aspect described above as a standard substance. The sixth aspect of the present invention consists in providing a reagent kit for the determination of denatured lipoprotein containing stabilized denatured lipoprotein produced in accordance with the first or the third aspect described above as a standard substance.

The method for determining the denatured lipoprotein in the fifth aspect described above does not need to be particularly restricted but may be selected from among the methods which have been heretofore known as useful for the purpose. As concrete examples of the method, such methods as radioimmunoassay (RIA), enzyme immunoassay (ELISA), fluorimmunoassay (FIA), luminescent immunoassay, agglutination immunoassay, immunonephelometry, and nephelometric immunoassay which are immunologic determination of denatured lipoprotein contained in a blood component by causing a given sample to contact the lipoprotein to an antibody capable of recognizing the denatured lipoprotein and measuring the reactivity of the antibody with the sample may be cited. As the measuring method, competitive assay and sandwich technique may be cited. Among the methods enumerated above, such methods as radioimmuno-assay, enzyme immunoassay, fluorescent immunoassay, and luminescent

immunoassay which effect the determination immunologically permit particularly favorable use of the stabilized denatured lipoprotein according to the present invention as a standard substance.

5 In the sixth aspect of the invention mentioned above, the reagent kit for the determination of denatured lipoprotein does not need to be particularly restricted but is only required to contain stabilized denatured lipoprotein according to the present invention as a standard substance. This reagent kit
10 has the same structure as the known kit, except it uses stabilized denatured lipoprotein according to the present invention as a standard substance. When the stabilized denatured lipoprotein contemplated by the present invention is used as a standard substance in the ELISA method, for example,
15 this reagent kit comprises a diluting liquid for a sample, a solid phase formed by immobilizing an antibody, a reaction buffer, a washing solution, a labeled secondary antibody (preferably an enzyme labeled secondary antibody), a detecting reagent (for example, a coloring fluid), and the
20 whole or part of the stabilized denatured lipoprotein according to the present invention as a standard substance. The mode of embodiment just described is also embraced in the concept of the present invention. The seventh aspect of the present invention, therefore, consists in providing a
25 reagent kit for the determination of denatured lipoprotein which comprises a diluting liquid for sample, a solid phase formed by immobilizing an antibody, a reaction buffer, a washing solution, a labeled secondary antibody, a detecting reagent, and the whole or part of stabilized denatured
30 lipoprotein produced as a standard substance according to the first or third aspect mentioned above as component elements.

In the seventh aspect of this invention described above, when the standard substance is not contained in the reagent kit but is still retained on the precondition that it will be substantially used in a kit, the standard substance
5 according to the present invention ought to be recognized as a component element for the reagent kit.

Now, the method for the production of denatured lipoprotein according to the present invention and the effect thereof will be described more specifically below with
10 reference to the following working examples resorting to the ELISA method for the determination of the activity of denatured lipoprotein. Of course, the present invention does not need to be limited to the following working examples.

Example 1

(1) Preparation of LDL

From the human blood plasma obtained by using EDTA as an anticoagulant, a part having specific gravity in the range of 1.019 - 1.063 was collected as an LDL fraction by the ultracentrifugation technique (by adjusting a given sample
20 to a specific gravity of 1.019 at 10°C, centrifuging the sample with 120,000×g for 20 hours, collecting the supernatant formed consequently, adjusting the supernatant to a specific gravity of 1.063, and further centrifuging it with 120,000×g for 24 hours). In this case, the purity of the LDL was confirmed
25 by the fact that the sample formed a single sharp band when it was assayed by the agarose electrophoresis technique.

Then, this LDL fraction was purified by dialyzing thoroughly for 16 hours or overnight against PBS (pH 7.4) containing 0.25 mmol/L of EDTA. The LDL purified as described
30 above was dissolved in PBS (pH 7.4) containing 0.25 mmol/L of EDTA until an LDL protein concentration reached 1 mg/mL and preserved in the dissolved state at 4°C until it was put

to use. In the present example, the mass of protein was determined by the Lowry modified Method. To be precise, this determination was effected by preparing a reagent by mixing a solution containing 2 (w/v) % of sodium carbonate, 0.4 (w/v) % of sodium hydroxide, 0.16 (w/v) % of tartaric acid, 1 (w/v) % of SDS and a solution containing 4 (w/v) % copper sulfate at a ratio of 100 : 1, mixing this reagent of 1.5 mL respectively with a given samples or a standard substance (BSA) of 0.5 mL in volume, allowing the relevant components to react at room temperature for 20 minutes, immediately mixing the resultant reaction solution with 0.15 mL of phenol reagent, allowing them to react at room temperature for 45 minutes, and measuring the produced reaction for absorbance at 660 nm.

15 (2) Oxidation of LDL

The LDL prepared in (1) was deprived of EDTA by dialyzing not less than 3 times (for not less than two hours/each) against not less than 500-1000 times as large in volume of PBS (pH 7.4) containing no EDTA and then dissolved in PBS (pH 7.4) containing no EDTA until the concentration of LDL reached 200 $\mu\text{g/mL}$. Then, 10 mL of the resultant LDL solution and copper sulfate (CuSO_4) added thereto until the final concentration reached 5 $\mu\text{mol/L}$ were left incubating at 37°C for three hours to effect oxidation of the LDL. To add EDTA to the solution until the final concentration of EDTA reached 1 mmol/L, the oxidation was stopped. The solution was deprived of CuSO_4 by dialyzing not less than three times (for not less than two hours/each) against not less than 500-1000 times as large in volume of PBS (pH 7.4) containing 1 mmol/L of EDTA. The oxidized LDL consequently prepared was preserved at 4°C.

30 In the present working example, the expression of the amount of the oxidized LDL was defined by the mass of protein

of the LDL as the raw material.

(3) Preparation of freeze-dried product of oxidized LDL

The oxidized LDL prepared in (2) was diluted with PBS (pH 7.4) until the concentration of protein reached 6.25 ng/mL in one portion (referred to as "L type") and 12.5 ng/mL in another portion (referred to as "H type"). The two types were each mixed well with BSA added thereto until the final concentration reached 2 (w/v) % and sucrose was added thereto until the final concentration reached 5 (w/v) % respectively. Then, the produced mixed solutions were each dispensed in a unit volume of 1 mL into glass vials, frozen by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.) at -50°C for 16 hours, freeze-dried at 20°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, then stoppered, and preserved at 4°C. At this time, each of the water contents of the freeze-dried products was 0.8 mass %.

(4) Formation of calibration curve with the freeze-dried product of oxidized LDL as standard substance

a. Production of peroxidase labeled antibody

One (1) mL of purified anti-human apo-B antibody (goat, Nippon Chemi-con Corp.) solution (5 mg/mL in concentration in 0.1 mol/L of borate buffer, pH 8.0) and 50 µL of 2-iodinothiolan-HCl solution (60 mmol/L in concentration in 0.1 mol/L of borate buffer, pH 8.0) added thereto were left reacting with each other at 30°C for 30 minutes. The resultant reaction solution was treated with a sephadex G-25 column (1 cm × 30 cm) (Pharmacia Corp.) equilibrated with 0.1 mol/L of phosphate buffer (pH 6.0) containing 5 mmol/L of EDTA. The antibody fraction eluted from the column was collected. One (1) mL of horseradish peroxidase (hereinafter abbreviated as "HRP," Toyobo K.K.) Solution (10 mg/mL in concentration

in 0.1 mol/L of phosphate buffer, pH 7.0) and 50 μ l of EMCS solution (50 mmol/L of DMSO solution, Dojin Kagakusha K.K.) added thereto were left reacting with each other at 30°C for 30 minutes. The resultant reaction solution was treated with a sephadex G-25 column (1 cm \times 30 cm) (Pharmarcia Corp.) equilibrated with 0.1 mol/L of phosphate buffer (pH 6.5). The HRP fraction eluted from the column was collected. The antibody fraction and the HRP fraction collected as described above were mixed and left reacting with each other at 30°C for 30 minutes. The resultant reaction solution was treated with a sephadex G-200 column (1 cm \times 100 cm) (Pharmarcia Corp.) equilibrated with 0.1 mol/L of phosphate buffer (pH 7.0). The antibody-HRP conjugate fractions eluted from the column were mixed and collected. The mass consequently collected was designated as "peroxidase-labelled anti-human apo-B antibody." The collected fraction was immediately diluted with BSA until a final concentration reached 1 (w/v) % and preserved in the diluted state at -50°C until use.

b. Preparation of DLH3 antibody

Male Balb/c mice not less than 8 weeks old, after administration by intra-abdominal injection of pristane (2,6,10,14-tetramethylpentadecane) with a dose of 0.5 mL/individual were each reared for two weeks. Then, the mice were each administrated to intra-abdominally with a hybridoma cell line FOH1a/DLH3, a cell capable of yielding a required monoclonal antibody (Deposit No. FERM BP-7171; J. Biol. Chem. 1994, 269: 15274-15279; and Patent publication JP-A-07-238,098) with a dose of 1×10^6 /individual. 7-14 days later, when the mice each accumulated ascites amply in the abdomen, the ascites was collected from the abdomen by the use of an 18G syringe and centrifuged at 3000 rpm for 10 minutes. The supernatant consequently formed was collected. To this

supernatant, an equal amount of PBS (pH 7.4) was added. To the produced mixture, a saturated ammonium sulfate solution equal in amount to the mixture was added dropwise over a period of one hour as kept thoroughly stirred. The resultant mixture
5 was continuously stirred further for one hour and then centrifuged at 3000 rpm for 10 minutes to discard the supernatant and collect the sediment. Further, the sediment was dissolved in PBS (pH 7.4) containing 0.5 mol/L of NaCl. The produced solution was treated with a Sephacryl S-300 column
10 (2.5 cm x 100 cm) (Pharmacia Corp.) equilibrated with PBS (pH 7.4) containing 0.5 mol/L of NaCl to collect a IgM fraction, which was designated as "DLH3 antibody." The concentration of the DLH3 antibody (mg/mL) was determined by measuring a given sample for absorbance at 280 nm in a light path 1 cm
15 in length and dividing the obtained absorbance by 1.3.

c. Sandwich ELISA analysis

The freeze-dried product of the oxidized LDL prepared in (3) was dissolved in 1 mL of purified water and then diluted with PBS containing 1 (w/v) % of BSA till varying stated
20 concentrations (0 ng/mL, 3.125 ng/mL, 6.25 ng/mL, 12.5 ng/mL, and 25 ng/mL).

In the individual wells of a 96F microplate (NALGE NUNC International K.K.), the DLH3 antibody prepared in b. above and diluted with Tris-HCl (pH 8.0) to 10 µg/mL was dispensed
25 in a unit amount of 1 µg/well and incubated at 4°C for 16 hours. The antibody solution formed in the wells was discarded. The residue in each of the wells was blocked by being incubated together with 350 µL of Tris-HCl (pH 8.0) containing 1 (w/v) % of BSA at room temperature for two hours. The blocked antibody
30 solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20.

The diluted solution containing the freeze-dried product

of oxidized LDL of a prescribed concentration prepared as described above was dispensed in a unit volume of 100 μ L to the individual wells, incubated at room temperature for two hours, and then washed four times with PBS (pH 7.4) containing
5 0.05 (v/v) % of Tween20.

To the individual wells, 100 μ L of the solution obtained by diluting the peroxidase labeled anti-human apo-B antibody prepared in "a." above with PBS (pH 7.4) containing 1 (w/v) % of BSA to a volume of 1000 times the original volume was placed
10 and incubated at room temperature for 30 minutes. The incubated solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20 and then left tinting for 30 minutes with 100 μ L of 0.03 (w/v) % of an aqueous hydrogen peroxide solution containing 3 mg/mL of o-phenylenediamine
15 (Wako Pure Chemical Industries Ltd.). The reaction still continuing in the solution was stopped by the addition of 50 μ L of 1 mol/L sulfuric acid. The solution was then measured absorbance at 492 nm. The results are shown in Fig. 1. By the use of the freeze-dried product of oxide LDL of the present
20 invention, a fine calibration curve could be formed as illustrated in Fig. 1.

(5) Comparison of stability of preservation

The freeze-dried products of the oxidized LDL prepared in (3) [standard oxidized LDL (1) (H type), 12.5 ng and standard
25 oxidized LDL (2) (L type), 6.25 ng] were preserved at 4°C for stated periods (0, 1, 3, and 4 weeks). The oxidized LDL prepared in (2) was diluted with PBS (pH 7.4) to two protein concentrations of 6.25 ng/mL (hereinafter referred to as "L type") and 12.5 ng/mL (hereinafter referred to as "H type").
30 BSA and sucrose were added to the two types to the final concentration of 2(w/v) % and 5 (w/v) % respectively, and the resultant solutions were mixed well. The resultant mixed

solutions were preserved in cold storage at 4°C without being freeze-dried [i.e. the oxidized LDL for comparison (1) (H type), 12.5 ng/mL and the oxidized LDL for comparison (2) (L type), 6.25 ng/mL] were preserved at 4°C for stated periods
5 (0, 1, 3, and 4 weeks).

After the preservation for the stated period, the standard oxidized LDLs (1) and (2) each dissolved in 1 mL of purified water and the oxidized LDLs for comparison (1) and (2) were treated by the same procedure as in (4) c. above
10 and measured absorbance at 492 nm. The results are shown in Fig. 2.

As shown in Fig. 2, the measurements obtained from the standard oxidized LDLs (1) and (2) freeze-dried in accordance with the present invention showed practically no change from
15 those obtained immediately after preparation (0 week), whereas the measurements obtained of the oxidized LDLs for comparison (1) and (2) after four weeks' preservation respectively showed falls of about 50% and about 45% from the measurements obtained immediately after preparation (0
20 week). These results clearly indicate that these oxidized LDLs for comparison were quite inferior in stability of preservation to the standard oxidized LDLs (1) and (2) conforming to the present invention.

Example 2

25 (1) Preparation of HDL

From the human blood plasma obtained by using EDTA as an anticoagulant, LDL was removed and a part having specific gravity in the range of 1.063 - 1.21 was collected as an HDL fraction by the ultracentrifugation technique (with 120,000×g
30 at 10°C for 48 hours). In this case, the purity of the HDL was confirmed by the fact that the sample formed a single sharp band when it was assayed by the agarose electrophoresis

technique.

Then, this HDL fraction was purified by dialyzing thoroughly for 16 hours against PBS (pH 7.4) containing 0.25 mmol/L of EDTA. The HDL purified as described above was dissolved in PBS (pH 7.4) containing 0.25 mmol/L of EDTA until an HDL protein concentration reached 1 mg/mL and preserved in the dissolved state at 4°C until it was put to use. In the present example, the mass of protein was determined by the Lowry modified Method described in Example 1

10 (2) Oxidation of HDL

The HDL prepared in (1) was deprived of EDTA by dialyzing not less than 3 times (for not less than two hours/each) against not less than 500-1000 times as large in volume of PBS (pH 7.4) containing no EDTA and then dissolved in PBS (pH 7.4) containing no EDTA until the concentration of HDL reached 100 µg/mL. Then, 10 mL of the resultant HDL solution and copper sulfate (CuSO₄) added thereto until the final concentration reached 10 µmol/L were left incubating at 37°C for 18 hours to effect oxidation of the HDL. To add EDTA to the solution until the final concentration of EDTA reached 1 mmol/L, the oxidation was stopped. The solution was deprived of CuSO₄ by dialyzing not less than three times (for not less than two hours/each) against not less than 500-1000 times as large in volume of PBS (pH 7.4) containing 1 mmol/L of EDTA. The oxidized HDL consequently prepared was preserved at 4°C.

In the present working example, the expression of the amount of the oxidized HDL was defined by the mass of protein of the HDL as the raw material.

30 (3) Preparation of freeze-dried product of oxidized HDL
BSA and sucrose were added to the oxidized HDL prepared in (2) until the final concentration reaches 2 (w/v) % and 5(w/v) % respectively, and the resultant solution was mixed

well. Then, the produced mixed solution was dispensed in a unit volume of 1 mL into glass vials, frozen by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.) at -50°C for 16 hours, freeze-dried at 5 20°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, then stoppered, and preserved at 4°C. At this time, each of the water contents of the freeze-dried products was 0.8 mass %.

(4) Formation of calibration curve with the freeze-dried product of oxidized HDL as standard substance

a. Adjustment of concentration

The freeze-dried product of the oxidized HCL prepared in (3) was dissolved in 1 mL of purified water added thereto and diluted with PBS containing 1 (w/v) % of BSA to varying 15 stated concentrations (0 ng/mL, 3.125 ng/mL, 6.25 ng/mL, 12.5 ng/mL, 25 ng/mL, 50 ng/mL, and 75 ng/mL).

b. Sandwich ELISA analysis

In the individual wells of a 96F microplate (NALGE NUNC International K.K.), an anti-human apo-AI mouse monoclonal 20 antibody (Nippon Chemi-con Corp.) solution diluted with a carbonate buffer (pH 9.5) to a concentration of 10 µg/mL was dispensed in a unit volume of 0.1 mL/well and incubated at 4°C for 16 hours. The antibody solution formed in the wells was discarded. The residue in each of the wells was blocked 25 by being incubated together with 350 µL of PBS (pH 7.4) containing 1 (w/v) % of BSA at room temperature for two hours. The blocked antibody solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20.

The diluted solution containing the freeze-dried product 30 of oxidized HDL of a prescribed concentration prepared as described in "a." above was dispensed in a unit volume of 100 µL to the individual wells, incubated at room temperature

for two hours, and then washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20.

To the individual wells, a DLH3 antibody diluted with PBS (pH 7.4) containing 1 (w/v) % of BSA was dispensed in
5 a unit volume of 100 μ L and incubated at room temperature for one hour. The incubated solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20 and then incubated together with 100 μ L of a peroxidase labeled anti-mouse IgM antibody (Zymed Laboratories, Inc.) diluted
10 to 1000 times with PBS containing 1 (w/v) % BSA at room temperature for 30 minutes. The resultant incubated solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20 and then left tinting with 100 μ L of 0.03 (w/v) % of an aqueous hydrogen peroxide solution containing 3 mg/mL
15 of o-phenylenediamine. The reaction still continuing in the solution was stopped by the addition of 50 μ L of 1 mol/L sulfuric acid. The solution was then measured absorbance at 492 nm. The results are shown in Fig. 3. By the use of the freeze-dried product of oxide HDL of the present invention, a fine
20 calibration curve could be formed as illustrated in Fig. 3.

(5) Comparison of stability of preservation

The freeze-dried products of the oxidized HDL prepared in (3) [standard oxidized HDL (1) (H type), 12.5 ng and standard oxidized HDL (2) (L type), 6.25 ng] were preserved at 4°C for
25 stated periods (0, 1, 2, 3, and 4 weeks). The oxidized HDL prepared in (2) was diluted with PBS (pH 7.4) to two protein concentrations of 6.25 ng/mL (hereinafter referred to as "L type") and 12.5 ng/mL (hereinafter referred to as "H type"). BSA and sucrose were added to the two types to the final
30 concentration of 2(w/v)% and 5(w/v)% respectively, and the resultant solutions were mixed well. The resultant mixed solutions were preserved in cold storage at 4°C without being

freeze-dried [i.e. the oxidized HDL for comparison (1) (H type), 12.5 ng/mL and the oxidized HDL for comparison (2) (L type), 6.25 ng/mL] were preserved at 4°C for stated periods (0, 1, 2, 3, and 4 weeks).

5 After the preservation for the stated period, the standard oxidized HDLs (1) and (2) each dissolved in 1 mL of purified water and the oxidized HDLs for comparison (1) and (2) were treated by the same procedure as in (4) b. above and measured absorbance at 492 nm. The results are shown in
10 Fig. 4.

As shown in Fig. 4, the measurements obtained from the standard oxidized HDLs (1) and (2) freeze-dried in accordance with the present invention showed practically no change from those obtained immediately after preparation (0 week),
15 whereas the measurements obtained of the oxidized HDLs for comparison (1) and (2) after cold storage respectively showed falls of about 40% and about 30% from the measurements obtained immediately after preparation (0 week). These results clearly indicate that these oxidized HDLs for comparison were
20 quite inferior in stability of preservation to the standard oxidized HDLs (1) and (2) conforming to the present invention.

Example 3

(1) Preparation of Lp(a)

From the human blood plasma obtained by using EDTA as
25 an anticoagulant, a part having specific gravity in the range of 1.060 - 1.125 was collected and further subjected to gel filtration with Biogel A-5m (Bio-rad Corp.) to collect a Lp(a) fraction. In this case, the purity of the Lp(a) was confirmed by the fact that the sample formed a single sharp band when
30 it was assayed by the agarose electrophoresis technique.

Then, this Lp(a) fraction was purified by dialyzing thoroughly (for 16 hours) against PBS (pH 7.4) containing

0.25 mmol/L of EDTA. Further, the Lp(a) purified as described above was dissolved in PBS (pH 7.4) containing 0.25 mmol/L of EDTA until an Lp(a) protein concentration reached 1 mg/mL and preserved in the dissolved state at 4°C until it was put
 5 to use. In the present example, the mass of protein was determined by the Lowry modified Method.

(2) Oxidation of Lp(a)

The Lp(a) prepared in (1) was deprived of EDTA by dialyzing not less than 3 times (for not less than two hours/each) against
 10 not less than 500-1000 times as large in volume of PBS (pH 7.4) containing no EDTA and then dissolved in PBS (pH 7.4) containing no EDTA until the concentration of Lp(a) reached 100 µg/mL. Then, 10 mL of the resultant LDL solution and copper sulfate (CuSO₄) added thereto until the final concentration
 15 reached 10 µmol/L were together left incubating at 37°C for 18 hours to effect oxidation of the Lp(a). To add EDTA to the solution until the final concentration of EDTA reached 1 mmol/L, the oxidation was stopped. The solution was deprived of CuSO₄ by dialyzing 2-3 times (for not less than two
 20 hours/each) against not less than 500-1000 times as large in volume of PBS (pH 7.4) containing 1 mmol/L of EDTA. The oxidized Lp(a) consequently prepared was preserved at 4°C.

In the present working example, the expression of the amount of the oxidized Lp(a) was defined by the mass of protein
 25 of the Lp(a) as the raw material.

(3) Preparation of freeze-dried product of oxidized Lp(a)

BSA and sucrose were added to the oxidized Lp(a) prepared in (2) until the final concentration reaches 2 (w/v) % and
 30 5 (w/v) % respectively, and the resultant solution was mixed well. The resultant mixture was dispensed in a unit volume of 1 mL into glass vials, then frozen at -50°C for 16 hours

by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.), freeze-dried at 20°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, then stoppered, and preserved
5 at 4°C. At this time, each of the water contents of the freeze-dried products was 0.8 mass %.

(4) Formation of calibration curve with the freeze-dried product of oxidized Lp(a) as standard substance

a. Adjustment of concentration

10 The freeze-dried product of the oxidized Lp(a) prepared in (3) was dissolved in 1 mL of purified water to varying stated concentrations (0 ng/mL, 0.15625 ng/mL, 0.3125 ng/mL, 0.625 ng/mL, 1.25 ng/mL, 2.5 ng/mL, and 5 ng/mL).

b. Sandwich ELISA analysis

15 In the individual wells of a 96F microplate (NALGE NUNC International K.K.), an anti-human Lp(a) mouse monoclonal antibody (Nippon Chemi-con Corp.) solution diluted with a carbonate buffer (pH 9.5) to a concentration of 10 µg/mL was dispensed in a unit amount of 1 µg/well and incubated at 4°C
20 for 16 hours. The antibody solution formed in the wells was discarded. The residue in each of the wells was blocked by being incubated together with 350 µL of PBS (pH 7.4) containing 1 (w/v) % of BSA at room temperature for two hours. The blocked antibody solution was washed four times with PBS (pH 7.4)
25 containing 0.05 (v/v) % of Tween20.

The aqueous solution containing the freeze-dried product of the oxidized Lp(a) of a prescribed concentration prepared as described in "a." above was dispensed in a unit volume of 100 µL to the individual wells, incubated at room temperature
30 for two hours, and then washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20.

In the individual wells, the diluted solution obtained

by diluting the DLH3 antibody prepared in (4) b. of Example 1 to a concentration of 10 $\mu\text{g/mL}$ with a 20 mmol/L of Tris-HCl solution (pH 7.4) containing 1 (w/v) % of BSA was dispensed in a unit volume of 100 μL and incubated at room temperature
5 for one hour. Then, the incubated solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20 and then incubated together with 100 μL of a peroxidase labeled anti-mouse IgM antibody (Nippon Chemi-con Corp.) diluted to 1000 times with PBS containing 1 (w/v) % BSA at room temperature
10 for 30 minutes. The resultant incubated solution was washed four times with PBS (pH 7.4) containing 0.05 (v/v) % of Tween20 and then left tinting with 100 μL of 0.03 (w/v) % of an aqueous hydrogen peroxide solution containing 3 mg/mL of o-phenylenediamine. The reaction still continuing in the
15 solution was stopped by the addition of 50 μL of 1 mol/L sulfuric acid. The solution was then measured absorbance at 492 nm. The results are shown in Fig. 5. By the use of the freeze-dried product of oxide Lp(a) of the present invention, a fine calibration curve could be formed as illustrated in Fig. 5.

20 (5) Comparison of stability of preservation

The freeze-dried products of the oxidized Lp(a) prepared in (3) [standard oxidized Lp(a) (1) (H type), 12.5 ng and standard oxidized Lp(a) (2) (L type), 6.25 ng] were preserved at 4°C for stated periods (0, 1, 2, 3, and 4 weeks). The oxidized
25 Lp(a) prepared in (2) was diluted with PBS (pH 7.4) to two protein concentrations of 6.25 ng/mL (hereinafter referred to as "L type") and 12.5 ng/mL (hereinafter referred to as "H type"). BSA and sucrose were added to the two types to the final concentration of 2(w/v)% and 5(w/v)% respectively,
30 and the resultant solutions were mixed well. The resultant mixed solutions were preserved in cold storage at 4°C without being freeze-dried [i.e. the oxidized Lp(a)s for comparison

(1) (H type), 12.5 ng/mL and the oxidized Lp(a) for comparison (2) (L type), 6.25 ng/mL] were preserved at 4°C for stated periods (0, 1, 2, 3, and 4 weeks).

After the preservation for the stated period, the standard oxidized Lp(a)s (1) and (2) each dissolved in 1 mL of purified water and the oxidized Lp(a)s for comparison (1) and (2) were treated by the same procedure as in "(4) c." above and measured absorbance at 492 nm. The results are shown in Fig. 6.

As shown in Fig. 6, the measurements obtained from the standard oxidized Lp(a)s (1) and (2) freeze-dried in accordance with the present invention showed practically no change from those obtained immediately after preparation (0 week), whereas the measurements obtained of the oxidized Lp(a)s for comparison (1) and (2) after four weeks' cold storage respectively showed falls of about 50% and about 40% from the measurements obtained immediately after preparation (0 week). These results clearly indicate that these oxidized Lp(a)s for comparison were quite inferior in stability of preservation to the standard oxidized Lp(a)s (1) and (2) conforming to the present invention.

Example 4

(1) Preparation of freeze-denatured human blood plasma
Blood samples were drawn from four subjects with heparin as an anticoagulant. Blood plasma samples were collected respectively from the blood samples by the ordinary method and assayed by a method similar to the ELISA method described in Example 1 (4).

Then, the blood plasma samples were each dispensed in a unit volume of 2 mL into glass vials, which were transferred from room temperature (25°C) to a freezer having an inner temperature of -30°C, left standing therein for not less than

three hours, returned to room temperature, and completely melted the whole contents. The individual blood plasma that had undergone the process including the freezing operation was analyzed by a procedure similar to the ELISA method. 5 described in Example 1 (4). The results are shown in Fig. 7. Fig. 7 clearly indicates that when the process including the freezing operation was performed on the blood plasma, the lipoprotein contained in the blood plasma was significantly denatured to produce a freeze-denatured 10 lipoprotein (oxidized LDL).

(2) Preparation of freeze-dried product of freeze-denatured human blood plasma

The blood plasma samples that had undergone the process including total 4 times of the freezing operation described 15 in (1) above were mixed in respectively equal volumes. The denatured LDL (oxidized LDL) concentration of the resultant mixed solution was calculated from the calibration curve formed as illustrated in Example 1 (4) in accordance with the ELISA method described in Example 1 (4).

20 Then, the freeze-denatured human blood plasma prepared in (1) was diluted to two oxidized LDL concentrations, 6.25 ng/mL (L type) and 12.5 ng/mL (H type), with a 10 mmol/L phosphate buffer (pH 7.4) containing 140 mmol/L of NaCl, BSA of final concentration of 2 (w/v) %, sucrose of final 25 concentration of 5 (w/v) %, and 0.25 mmol/L of EDTA-2Na. The two types of diluted solution were dispensed in a unit volume of 1 mL into glass vials, frozen at -50°C for 16 hours by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.), then freeze-dried at 5°C under 30 a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, stoppered, and preserved at 4°C until use. This was designated as "freeze-dried product of

freeze-denatured human blood plasma." Each of the water contents of the freeze-dried products was 0.8 mass %.

(3) Determination of concentration of freeze-dried product of freeze-denatured human blood plasma

5 The freeze-dried product of the freeze-denatured human blood plasma prepared in (2) was dissolved in 1 mL of purified water added thereto. The resultant solution was measured absorbance at 492 nm in accordance with the method for determining the oxidized LDL by the sandwich ELISA method
10 described in Example 1 (4). When the oxidized LDL concentration of the solution was sought from the calibration curve formed in Example 1 (4) based on the absorbance found by the measurement, the oxidized LDL concentration in the freeze-dried product showed practically no discernible change
15 before and after the step of freeze-drying.

(4) Comparison of stability of preservation

 The two types of freeze-dried product [standard freeze-denatured human blood plasma (1) (H type), 12.5 ng and standard freeze-denatured human blood plasma (2) (L type),
20 6.25 ng] of the freeze-denatured human blood plasma prepared in (2) were preserved at 4°C for varying stated periods (0, 6, and 12 months). Then, the two types of freeze-dried product of oxidized LDL [standard oxidized LDL (1) (H type), 12.5 ng and standard oxidized LDL (2) (L type), 6.25 ng] prepared
25 in Example 1 (3) were preserved at 4°C for varying stated periods (0, 6, and 12 months).

 After the preservation for the stated period, the standard freeze-denatured human blood plasmas (1) and (2) and the standard oxidized LDLs (1) and (2) were each dissolved
30 in 1 mL of purified water. The resultant solutions were assayed by following a procedure similar to the ELISA method described in Example 1 (4). The results are shown in Table

1 below.

Table 1

Duration of preservation (months)	OD ₄₉₂			
	Standard freeze- denatured human blood plasma (1)	Standard freeze- denatured human blood plasma (2)	Standard oxidized LDL (1)	Standard oxidized LDL (2)
0	0.740	0.272	0.758	0.283
6	0.727	0.270	0.508	0.222
12	0.677	0.274	0.430	0.207

5 As shown in Table 1, the standard oxidized LDL according to the present invention excelled in stability of preservation so long as it had a low concentration [standard oxidized LDL (2)]. The standard freeze-denatured human blood plasma according to the present invention which attained
10 denaturation of lipoprotein by freezing and then acquired stability by freeze-drying significantly excelled compared with the standard oxidized LDL of the present invention in terms of stability of prolonged preservation even when it had a high concentration [standard freeze-denatured human
15 blood plasma (1)] compared with the standard oxidized LDL of the present invention.

Example 5

(1) Preparation of freeze-denatured human blood serum
Blood samples were drawn from four subjects. Blood serum
20 samples were separated respectively from the blood samples by the ordinary method and mixed. BSA and sucrose were added to the blood serum to the final concentration of 2(w/v)% and

5(w/v)% respectively, and the resultant mixtures were mixed well. The mixtures were dispensed in a unit volume of 2 mL into glass vials, frozen by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.) at -50°C for 16 hours, freeze-dried at 5°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, stoppered, and preserved at 4°C. Each of the water contents of the freeze-dried products was 0.8 mass %.

(2) Preparation of freeze-dried product of freeze-denatured human blood serum

Purified water was added in a unit volume of 2 mL to the glass vials containing the freeze-denatured human blood plasma samples prepared in (1), thereby dissolved the contents of the vials thoroughly. The oxidized LDL concentrations of the produced solutions were calculated from the calibration curve formed as illustrated in Example 1 (4) in accordance with the ELISA method described in Example 1 (4).

Then, the freeze-denatured human blood serum prepared in (1) was diluted to two oxidized LDL concentrations, 6.25 ng/mL (L type) and 12.5 ng/mL (H type), with a 10 mmol/L phosphate buffer (pH 7.4) containing 140 mmol/L of NaCl, BSA of final concentration of 2 (w/v) %, sucrose of final concentration of 5 (w/v) %, and 0.25 mmol/L of EDTA-2Na. The two types of diluted solution were dispensed in a unit volume of 1 mL into glass vials, frozen at -50°C for 16 hours by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.), then freeze-dried at 5°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, stoppered, and preserved at 4°C until use. This was designated as "freeze-dried product of freeze-denatured human blood plasma." Each of the water contents of the freeze-dried products was 0.8 mass %.

(3) Determination of concentration of freeze-dried product of freeze-denatured human blood serum

The freeze-dried product of the freeze-denatured human blood serum prepared in (2) was dissolved in 1 mL of purified water added thereto. When the oxidized LDL concentration of the resultant solution was sought from the calibration curve formed in Example 1 (4) by the method for determination of the oxidized LDL in accordance with the ELISA method described in Example 1 (4), the oxidized LDL concentration in the freeze-dried product showed practically no discernible change before and after the step of freeze-drying.

(4) Comparison of stability of preservation

The freeze-dried product of the freeze-denatured human blood serum prepared in (2) was dissolved with 1 mL of purified water into two oxidized LDL concentrations of 12.5 ng/mL and 6.25 ng/mL respectively to prepare the standard freeze-denatured human blood serum samples, (1) (H type) and (2) (L type). These two types of samples were preserved at 4°C for varying stated periods (0, 3, 7, 10, and 14 days). Then, the two types of freeze-dried product of the oxidized LDL prepared in Example 1 (3) were dissolved in the same manner as described above to obtain standard oxidized LDLs having oxidized LDL concentrations of 12.5 ng/mL and 6.25 ng/mL i.e., standard oxidized LDL (3) (H type) and standard oxidized LDL (4) (L type). These types were preserved in a dissolved state at 4°C for varying stated periods (0, 3, 7, 10, and 14 days).

After the preservation for the stated period, the standard freeze-denatured human blood plasmas (1) and (2) and the standard oxidized LDLs (3) and (4) were each assayed by following the method for determination of oxidized LDL in accordance with the sandwich ELISA method described in Example 1 (4). The results are shown in Table 2 and Fig. 8.

Table 2

Duration of preservation (days)	OD ₄₉₂			
	Standard freeze- denatured human blood serum (1)	Standard freeze- denatured human blood serum (2)	Standard oxidized LDL (3)	Standard oxidized LDL (4)
0	0.8175	0.3693	0.7689	0.3608
3	0.8107	0.3533	0.6762	0.3615
7	0.7602	0.3387	0.6705	0.3473
10	0.7800	0.3440	0.5772	0.3240
14	0.7630	0.3400	0.5760	0.2985

As shown in Table 2, the standard oxidized LDL according to the present invention excelled in stability of preservation so long as it had a low concentration [standard oxidized LDL (4)]. The standard freeze-denatured human blood serum according to the present invention which attained denaturation of lipoprotein by freezing and then acquired stability by freeze-drying significantly excelled compared with the standard oxidized LDL of the present invention in terms of stability of prolonged preservation even when it had a high concentration [standard freeze-denatured human blood serum (1)].

Example 6

(1) Preparation of freeze-denatured LDL

The LDL prepared in Example 1 (1) was deprived of EDTA by dialyzing not less than three times (not less than two hours/each) against not less than 500 - 1000 times as large

in volume of PBS (pH 7.4) containing no EDTA, diluted to a LDL concentration of 1 mg/mL with a 10 mmol/L phosphate buffer (pH 7.4) containing 140 mmol/L of NaCl, BSA of a final concentration of 2 (w/v) %, sucrose of a final concentration of 5 (w/v) %, and 0.25 mmol/L of EDTA-2Na, and mixed well. Then, the produced diluted solution was dispensed in a unit volume of 2 mL into glass vials, which were transferred from room temperature (22°C) to freezers having inner temperatures of -85°C and -30°C respectively, left standing therein for not less than one hour, and returned to room temperature to effect thorough dissolution of the contents of the vials. The process including the step of freezing and the dissolving mentioned above was performed up to a total of 10 repetitions. The contents of the glass vials were partly collected during some intervals between the said processes including the steps of freezing and melting. The samples thus obtained were assayed by the sandwich ELISA method described in Example 1 (4). The results are shown in Fig. 9.

As shown in Fig. 9, the denaturation of LDL nearly reached a saturated state when the process including the steps of freezing and melting was performed up to three repetitions while the internal temperature of the freezer was -30°C. When the internal temperature of the freezer was -85°C, however, the denaturation of LDL did not reach a saturated state unless the process including the steps of freezing and melting was performed up to nine repetitions. The results indicate that the product amount of denatured LDL to be produced varies with the lowered temperature.

(2) Preparation of freeze-dried product of freeze-denatured LDL

The freeze-denatured LDL prepared by performing the process including the step of freezing at the internal

temperature, -30°C , of the freezer in (1) up to a total of four repetitions was diluted to oxidized LDL concentrations, 6.25 ng/mL (L type) and 12.5 ng/mL (H type), with 10 mmol/L phosphate buffer (pH 7.4) containing 140 mmol/L of NaCl, BSA of final concentration of 2 (w/v) %, sucrose of final concentration of 5 (w/v) %, and 0.25 mmol/L of EDTA-2Na. The two types of diluted solution thus obtained were dispensed in a unit volume of 1 mL into glass vials, frozen at -50°C for 16 hours by the use of a vacuum freeze-drying device, Kyowa Drier RL-201BS (Kyowa Shinku Gijutsu K.K.), then freeze-dried at 5°C under a reduced pressure of 1.33 Pa for 48 hours to gasify and expel the water component, stoppered, and preserved at 4°C until use. This was designated as "freeze-dried product of freeze-denatured LDL. Each of the water contents of the freeze-dried products was 0.8 mass %.

(3) Determination of concentration of freeze-dried product of freeze-denatured LDL

The freeze-dried product of the freeze-denatured LDL prepared in (2) was dissolved in 1 mL of purified water added thereto. When the oxidized LDL concentration of the resultant solution was sought from the calibration curve formed in Example 1 (4) by the method for determination of the oxidized LDL in accordance with the ELISA method described in Example 1 (4), the oxidized LDL concentration in the freeze-dried product showed practically no discernible change before and after the step of freeze-drying.

(4) Comparison of stability of preservation after dissolution

The freeze-dried product of the freeze-denatured LDL prepared in (2) was dissolved with 1 mL of purified water into two oxidized LDL concentrations, 12.5 ng/mL and 6.25 ng/mL, to prepare the standard freeze-denatured human LDLs,

(1) (H type) and (2) (L type) respectively. These two types of samples were preserved at 4°C for varying stated periods (0, 3, 7, 10, and 14 days). Then, the two types of freeze-dried product of the oxidized LDL prepared in Example 1 (3) were dissolved in the same manner as described above to obtain standard oxidized LDLs having oxidized LDL concentrations of 12.5 ng/mL and 6.25 ng/mL [standard oxidized LDL (3) (H type) and standard oxidized LDL (4) (L type)]. These types were preserved in a dissolved state at 4°C for varying stated periods (0, 3, 7, 10, and 14 days)

After the preservation for the stated period, the standard freeze-denatured LDLs (1) and (2) and the standard oxidized LDLs (3) and (4) were each measured absorbance at 492 nm by following the method for determination of oxidized LDL in accordance with the sandwich ELISA method described in Example 1 (4). The oxidized LDL contents in the standard samples were sought from the calibration curve formed in Example 1 (4), based on the absorbance obtained by the measurement above. The results are shown in Table 3 and Fig. 10.

Table 3

Duration of preservation (days)	OD ₄₉₂			
	Standard freeze-denatured LDL (1)	Standard freeze-denatured LDL (2)	Standard oxidized LDL (3)	Standard oxidized LDL (4)
0	0.8633	0.3826	0.7689	0.3608
3	0.8265	0.3532	0.6762	0.3615
7	0.7924	0.3516	0.6705	0.3473
10	0.7952	0.3439	0.5772	0.3240
14	0.7886	0.3476	0.5760	0.2985

As shown in Table 3, the standard oxidized LDL according to the present invention excelled in stability of preservation so long as it had a low concentration [standard oxidized LDL (4)]. The standard freeze-denatured human blood serum according to the present invention which attained denaturation of lipoprotein by freezing and then acquired stability by freeze-drying significantly excelled compared with the standard oxidized LDL of the present invention in terms of stability of prolonged preservation after dissolution even when it had a high concentration [standard freeze-denatured LDL (1)].

Industrial Applicability of the Invention

The present invention, as described above, concerns denatured lipoprotein possessed of excellent stability of preservation which is produced by freeze-drying denatured lipoprotein obtained by artificially denaturing lipoprotein and a method for the production thereof. The present invention further concerns a method for the production of

denatured lipoprotein which comprises a step of performing a process including at least one freezing operation on a solution containing denatured lipoprotein thereby denaturing the lipoprotein contained in the solution and denatured lipoprotein excelling in stability of preservation obtained by further freeze-drying the denatured lipoprotein obtained by the method mentioned above and a method for the production thereof. It is strongly suggested that the denatured lipoprotein is deeply related to various diseases of the circulatory system including such diseases of the coronary artery system as cardiac infarction and stenocardia, such diseases of the cerebral arteries as cerebral infarction and cerebrovascular dementia, such diseases of the renal arteries as nephropathy and diabetic nephropathy, and such diseases of the peripheral artery system as obstruction of peripheral arteries. The standard substance for the determination of denatured lipoprotein in blood and the reagent for the investigation of the physiological role and the physiological activity of denatured lipoprotein are very important substances which affect the results of such experiments.

By the method of the present invention, therefore, it has now become possible to produce denatured lipoprotein excelling in stability of preservation, to put it in other words, exhibiting definite determinations. In addition to the advantage mentioned above, the stabilized lipoprotein which is produced by freeze-drying denatured lipoprotein obtained by performing a process including at least one freezing operation on a solution containing denatured lipoprotein excels not only in stability of preservation but also in stability of preservation after dissolution. The stabilized denatured lipoprotein contemplated by the present invention retains the excellent stability even when it is

applied to the form of a solution, i.e. the forms as actual use. This fact makes this stabilized denatured lipoprotein highly advantageous for the sake of determining denatured lipoprotein.

5 The stabilized denatured lipoprotein according to the present invention, therefore, is useful as a standard substance in a method for determining denatured lipoprotein contained in a blood component by placing a given sample in contact with an antibody capable of recognizing denatured
10 lipoprotein and measuring the reactivity of this antibody on the sample and as a varying experimental reagent for investigating the physiological role and the physiological activity of denatured lipoprotein. Evidently, it exerts a very important effect on the commercialization of diagnostic
15 technology and the development of reagents with a view to various objects mentioned above.

CLAIMS

1. A method for producing stabilized denatured lipoprotein, comprising a step of artificially denaturing lipoprotein, thereby obtaining denatured lipoprotein and a
5 step of freeze-drying said denatured lipoprotein, thereby stabilizing said denatured lipoprotein.

2. A method according to claim 1, wherein said lipoprotein is human lipoprotein.

3. A method according to claim 1 or 2, wherein said
10 lipoprotein is at least one selected from the group consisting of chylomicrons, VLDL, LDL, Lp(a), HDL2, and HDL3.

4. A method according to any one of claims 1 - 3, wherein said denatured lipoprotein is obtained by oxidizing lipoprotein in the presence of a metal ion.

5. A method according to claims 4, wherein said metal
15 ion is a copper ion, an iron ion, or a mixture thereof.

6. A method according to any one of claims 1 - 3, wherein said denatured lipoprotein is obtained by acetylating lipoprotein.

7. A method according to any one of claims 1 - 3, wherein
20 said denatured lipoprotein is obtained by incorporating aldehyde in lipoprotein by the use of malondialdehyde.

8. A method according to any one of claims 1 - 7, which further comprises a step of adding a stabilizing agent.

9. A method according to claim 8, wherein said
25 stabilizing agent is at least one selected from the group consisting of sucrose, lactose, trehalose, bovine blood serum albumin (BSA), and human blood serum albumin (HSA).

10. A method for the production of denatured lipoprotein,
30 comprising a step of performing a process including at least one freezing operation on a solution containing lipoprotein thereby denaturing the lipoprotein contained in said

solution.

11. A method according to claim 10, wherein said denatured lipoprotein is used as a standard substance for determining denatured lipoprotein in blood or an experimental reagent for investigating the physiological role or the physiological activity of denatured lipoprotein.

12. A method according to claim 10 or 11, wherein said denatured lipoprotein is an oxidized lipoprotein or malondialdehyde-conjugated lipoprotein.

13. A method according to claim 10 or 11, wherein said denatured lipoprotein is capable of reacting with a DLH3 antibody which is yielded by hybridoma cell line, mouse-mouse hybridoma FOH1a/DLD3 (Deposit No. FERM BP-7171).

14. A method for producing stabilized denatured lipoprotein, comprising a step of performing a process including at least one freezing operation on a solution containing lipoprotein thereby denaturing the lipoprotein contained in said solution and obtaining denatured lipoprotein and a step of further freeze-drying said denatured lipoprotein thereby stabilizing said denatured lipoprotein.

15. Stabilized denatured lipoprotein produced by a method set forth in any one of claims 1 - 9 or in claim 14.

16. A method for the determination of denatured lipoprotein by using stabilized denatured lipoprotein set forth in claim 15 as a standard substance.

17. A method according to claim 16, wherein said stabilized denatured lipoprotein is used as a standard substance in a method for immunological determination.

18. A method according to claim 17, wherein said method for immunological determination is selected from among radio immunoassay, enzyme immunoassay, fluoroimmunoassay, luminescent immunoassay, agglutination immunoassay,

immunonephelometry, and nephelometric immunoassay.

19. A method according to claim 17, wherein said method for immunological determination is a competitive method or a sandwich method.

5 20. A reagent kit for the determination of denatured lipoprotein, comprising stabilized denatured lipoprotein set forth in claim 15 as a standard substance.

21. A reagent kit for the determination of denatured lipoprotein, comprising a diluting liquid for a sample, a
10 solid phase formed by immobilizing an antibody, a reaction buffer, a washing solution, a labeled secondary antibody, a detecting reagent, and the whole or part of stabilized denatured lipoprotein set forth in claim 15 as a standard substance as component elements.

15

ABSTRACT

Denatured lipoprotein excelling in stability of prolonged preservation (namely exhibiting only sparing liability to induce denaturation of lipoprotein) and used
5 as a standard substance for the determination of denatured lipoprotein in blood or the determination of the physiological activity of the lipoprotein and a method for the production thereof are provided. The method for the production of the stabilized denatured lipoprotein comprises artificially
10 denaturing lipoprotein, thereby obtaining denatured lipoprotein, freeze-drying the denatured lipoprotein or performing a process including at least one freezing operation on a solution containing denatured lipoprotein thereby denaturing the lipoprotein contained in the solution and
15 obtaining denatured lipoprotein and further freeze-drying the denatured lipoprotein.

COMBINED DECLARATION AND POWER OF ATTORNEY

10/009151

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and solo inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled STABILIZED DENATURED LIPOPROTEIN AND METHOD FOR PRODUCING THEREOF the specification of which

X is attached hereto
 was filed on November 30, 2001
 as Application Serial No. 10/009,151 and was amended on November 30, 2001
X was described and claimed in PCT International Application No. PCT/JP00/03413
 filed on May 26, 2000 and as amended under PCT Article 19, on _____

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United State of America filed by me on the same subject matter having a filing date before that of the application(s) on which priority is claimed:

COUNTRY	APPLICATION NO.	FILING DATE	PRIORITY CLAIMED
<u>Japan</u>	<u>11-155198</u>	<u>June 2, 1999</u>	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
_____	_____	_____	<input type="checkbox"/> Yes <input type="checkbox"/> No
_____	_____	_____	<input type="checkbox"/> Yes <input type="checkbox"/> No

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issued thereon.

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